Antibacterial, Cytotoxic and Antioxidant Activities of n-Hexane, Chloroform and Ethyl Acetate extracts of Trichosanthes cucumerina leaves

Israt Jahan Bulbul*, Khadiza-tul-Kubra, Mohita Chowdhury Shimu Department of Pharmacy, Southeast University Banani, Dhaka, Bangladesh * israt_jahanb872@yahoo.com



ABSTRACT

The main aim of this study was to find out the antibacterial, cytotoxic and antioxidant activities of n-hexane, chloroform and ethyl acetate extracts of *T. cucumerina* (Cucurbitaceae). Disc diffusion technique was used for *in vitro* antibacterial screening against gram positive, gram negative human pathogenic bacteria. Here kanamycin disc (30 µg/disc) was used as standard. The chloroform and the n-hexane extract of *T. cucumerina* showed moderate antibacterial activity with the average zone of inhibition 7-13 mm and 7-9 mm respectively. The brine shrimp lethality bioassay method was used to determine the cytotoxic activity and vincristine sulphate was used as positive control. Among the extractives the chloroform soluble fraction demonstrated the highest cytotoxic activity with LC₅₀ 17.09 µg/ml which indicates the compounds present in the chloroform extract are promisingly cytotoxic. Antioxidant activity test of the crude extracts were assessed by means of DPPH free radical scavenging method where ascorbic acid was used as standard. The ethyl acetate fraction of *T. cucumerina* showed strongest antioxidant activity with IC₅₀ value of 52.18 µg/ml. Incase of phenolic content, the n-hexane, chloroform and ethyl acetate extracts of *T. cucumerina* revealed 18.79, 31.33 and 29.04 mg of GAE / gm of extractives, respectively.

Key words: T. cucumerina, antibacterial, antioxidant, cytotoxic.

INTRODUCTION

Trichosanthes *cucumerina* var. anguina (L.) (Cucurbitaceae) is a tropical or subtropical vine, raised for its strikingly long fruit, used as a vegetable, medicine, and a lesser known use, crafting didgeridoos. Common names include snake gourd^[1]. T. cucumerina is highly constituted with proteins, fat, fiber, carbohydrates, vitamin A and E, total phenolics and flavonoids^[2]. The predominant mineral elements are potassium, phosphorus, sodium, magnesium and zinc ^[3]. The triterpenes found are 23, 24dihydrocucurbitacin D, 23, 24-dihydrocucurbitacin B, cucurbitacin B, 38-hydroxyolean- 13(18)-en-28-oic acid, 3-oxo-olean-13(18)-en-30-oic acid and the 3-*O*- β -D-glucopyranosyl-24 ξ -ethylcholest-7, sterol 22-dien-3 β -ol^[4]. α - carotene, β - carotene, ascorbic acid, lycopene are also found in it ^[5]. A novel isoflavone glucoside, 5,6,6'-trimethoxy-3',4'-methy lenedioxyisoflavone 7-O-beta-D-(2"-O-p coumaroyl glucopyranoside) has been characterized from the seeds of *Trichosanthes*^[6].

Decoctions of leaves and stems are used in the treatment of bilious disorders, skin diseases, cardiac tonic and emmenagogue. Ripe fruits possess purgative, anthelmintic and emetic properties. They improve appetite and cure biliousness. Seeds and root are used for the expulsion of intestinal worms and in the treatment of diarrhea and syphilis ^[7]. The petroleum ether extract of the seeds have been found to possess appreciable antibacterial activity ^[8]. Hot aqueous extract of root tubers of *T. cucumerina* have significant anti-inflammatory activity ^[9], the root and the fruit juice extract of *T. cucumerina* has cytotoxicity ^[10].

T. cucumerina showed significant blood glucose lowering activity ^[11] ^[12] ^[13], moderate larvicidal effects ^[14], good hepatoprotective activity ^[15], antihistamine activity ^[12], dose dependent gastroprotective effects ^[12].

How to cite this article: Bulbul IJ, Kubra KT, Shimu MC; Antibacterial, Cytotoxic and Antioxidant Activities of n-Hexane, Chloroform and Ethyl Acetate extracts of Trichosanthes cucumerina leaves; PharmaTutor; 2016; 4(1); 42-46

The objective of the present study was to investigate the antibacterial, cytotoxic and antioxidant activity of the different fractions of *T. cucumerina*. Therefore, systematic research with medicinal plants may open the door of many therapeutic choices.

MATERIALS AND METHODS

Plant material

The leaves of the plant *T. cucumerina* were collected during the month of July 2010 from the area of Moynertak, Tongi, Dhaka.

Plant materials extraction and fractionation

The fresh leaf was collected, sun dried for seven days and ground. The dried powder of *T. cucumerina* leaf (200 gm) was soaked in 600 ml of ethanol for 7 days and filtered through a cotton plug followed by Whatman filter paper number 1. The concentrated ethanolic extract of leaf was fractionated by the modified Kupchan partitioning method ^[16] into nhexane, chloroform and ethyl acetate. The subsequent evaporation of solvents afforded nhexane (450 mg), chloroform (700 mg) and ethyl acetate (350 mg) from leaf extract.

Antibacterial assay

In our present study, the antibacterial activity of nhexane, chloroform and ethyl acetate fractions of the plant were investigated in comparison with standard kanamycin (30 µg/ disc) against a number of pathogenic Gram-positive (Bacillus megaterium, Bacillus subtilis, Staphylococcus aureus and Sarcina lutea) and Gram- negative (Salmonella paratyphi, S. typhi, Vibrio parahaemolyticus, V. mimicus, Escherichia coli, Shigella dysenteriae, S. boydii and Pseudomonas aeruginosa) bacteria. The microorganisms were collected as pure cultures from the Institute of Nutrition and Food Science (INFS), University of Dhaka, Bangladesh. The sample solution of the material to be tested was prepared by dissolving a definite amount of material in methanol to attain a concentration of 50 mg/ml. 10 µl of such solution was applied on sterile disc (5 mm diameter, filter paper) and allowed to dry off the solvent in an aseptic hood. Thus, such discs contain 500 µg of crude extracts. To compare the activity with standard antibiotics, Kanamycin (30 µg/disc) was used.

CYTOTOXICITY SCREENING

Brine shrimp Lethality Bioassay

Brine shrimp lethality bioassay ^[17] ^[18] was used for probable cytotoxic activity. The eggs of Brine Shrimp (*Artemia salina*) was collected from local pet shops and hatched in a tank at a temperature around 37 °C with constant oxygen supply. Two days were allowed to hatch and mature the nauplii. Stock solution of the sample was prepared by dissolving required amount of extract in specific volume of pure dimethyl sulfoxide (DMSO) to attain concentrations of 5, 10, 20, 40 and 80 µg/ml. With the help of a pasteur pipette nauplii were exposed to different concentrations of the extracts.

DPPH radical scavenging activity

Antioxidant activity of n-hexane, chloroform and ethyl acetate of leaf extracts of *T. cucumerina* was determined on the basis of their scavenging potential of the stable DPPH free radical in both qualitative and quantitative assay.

Qualitative analysis

A suitably diluted stock solutions were spotted on precoated silica gel TLC plates and the plates were developed in solvent systems of different polarities (polar, medium polar and non-polar) to resolve polar and non-polar components of the extracts. The plates were dried at room temperature and were sprayed with 0.02% DPPH in methanol. Bleaching of DPPH by the resolved band was observed for 10 minutes and the color changes (yellow on purple background) were noted ^[19].

Quantitative analysis

The free radical scavenging activities (antioxidant capacity) of the plant extracts on the stable radical 1, 1-diphenyl-2- picrylhydrazyl (DPPH) were estimated by the method of Brand-Williams ^[20]. During this samples of n-hexane, the test experiment chloroform and ethyl acetate extracts of T. cucumering at different concentrations were mixed with 3.0 ml of DPPH methanol solution. The antioxidant potential was assayed from the bleaching of purple colored methanol solution of DPPH radical by the plant extracts as compared to that of ascorbic acid by UV spectrophotometer (UV-1501PC SHIMADZU, Japan) at 517 nm. Ascorbic acid was used as a positive control. Percent scavenging of the DPPH free radical was measured using the following equation-

% DPPH radical scavenging = [1 – (As/ Ac)] × 100

Here, Ac = absorbance of control, As = absorbance of sample solution.

Then % inhibitions were plotted against respective concentrations used and from the graph IC $_{50}$ was calculated. The lower IC $_{50}$ indicates higher radical scavenging activity and vice versa.

Assay for Total Phenolics

Total phenolic content of different parts of *T. cucumerina* extractives was measured employing the method as described by Skerget *et al.,* 2005 ^[21] involving Folin-Ciocalteu reagent as oxidizing agent and gallic acid as standard ^[22]. 0.5 ml of diluted plant extract and standard of different concentrations solution were taken in the test tube followed by adding 2.5 ml of Folin–ciocalteu (Diluted 10 fold with water) & 2 ml of Sodium carbonate (1 M) respectively. Solutions were then incubated for 20 minutes at 45°C in the water bath. The absorbance was measured colorimetrically at 760 nm to determine the total phenol contents by using standard curve prepared (Fig:1) from gallic acid solution with different concentration.

RESULTS AND DISCUSSION

Antibacterial activity

Different extractives of *T. cucumerina* were screened against human pathogenic organisms to evaluate antibacterial activities by disc diffusion method. The chloroform fraction possesses the zone of inhibition value ranged from 7-13 mm (Table: 1). Among different fractions tested, chloroform fraction of the plant exhibited moderate inhibitory activity followed by n-hexane fraction (7-9 mm) whereas ethyl acetate fraction showed little or no activity on the tested microorganisms. The most sensitivity was observed in *P. aeruginosa* (13 mm), *S. paratyphi* (11 mm) and *V. parahaemolyticus* (10 mm) by chloroform fraction of the plant.

Cytotoxicity screening

LC₅₀ value of chloroform, n-hexane and ethyl acetate fractions found with the value of 17.09 μ g/ml, 27.72 μ g/ml and 44.71 μ g/ml respectively in comparison with vincristine sulphate as standard whose LC₅₀ value 8.844 μ g/ml. Among them chloroform fraction of the plant exhibited the potent cytotoxic activity.

DPPH RADICAL SCAVENGING ACTIVITY

Qualitative assay

The color changes (yellow on purple background) on the TLC plates were observed due to the bleaching of DPPH by the resolved bands.

Quantitative assay

n-hexane, chloroform, ethyl acetate extracts of the plant showed significant antioxidant activity with the IC₅₀ value of 65.84 µg/ml, 59.01 µg/ml, 52.18µg/ml respectively compared with the standard ascorbic acid with IC₅₀ value of 45.47 µg/ml (Fig: 2), the fractions exhibited a concentration dependant DPPH radical scavenging activity.

Total phenolic content

The phenolic content of plant fractions was determined using the Folin–Ciocalteu assay and was expressed as gallic acid equivalents (GAE). The phenolic contents of n-Hexane, chloroform and ethyl acetate soluble fractions of *T. cucumerina* plant were 18.79 mg/g, 29.04 mg/g and 31.33 mg/g of the dry weight.

CONCLUSION

The present study indicates that the n-hexane, chloroform and ethyl acetate extracts of the different fractions of *T. cucumerina* exhibited mild to moderate antibacterial, profound antioxidant, total phenolic content and cytotoxic activities. The chloroform extract of the plant showed moderate antibacterial activity. So, the studied plant may have clinical and therapeutic proposition in the most life threaten diseases like tumor or cancer, various infectious diseases and the aging process of human being. Therefore, further investigation should be necessary for the development of novel lead compound.

PRINT ISSN: 2394-6679 | E-ISSN: 2347-7881

Test organism	Diameter of zone of inhibition			
	n-Hexane extract (500ug/disc)	Chloroform extract (500ug/disc)	Ethyl acetate extract (500ug/disc)	Kanamycin(30µg/ disc)
GRAM POSITIVE BACTERIA				
B. subtilis	8	8	0	24
B. megaterium	8	8	0	25
S. aureus	7	7	0	10
S. lutea	7	7	0	14
GRAM NEGATIVE BACTERIA				
V. mimicus	9	8	7	29
S. boydii	7	9	7	31
P. aeruginosa	9	13	0	32
S. typhi	8	8	0	32
S. paratyphi	8	11	0	29
V. parahaemolyticus	8	10	0	30
S. dysenteriae	0	7	0	9
E. coli	7	8	0	17





Figure 1: Determination of LC50 values for standard and crude n-hexane, chloroform and ethyl acetate extracts of leaves T. cucumerina from linear correlation between logarithms of concentration versus percentage of mortality.



Figure 2: Determination of IC_{50} value for standard and crude n-hexane, chloroform and ethyl acetate extracts of leaves of *T. cucumerina* from linear correlation between concentrations (μ g/ml) versus percentage of scavenging of DPPH.

↓ REFERENCES

1.Saboo SS, Thorat P, Tapadiya GG, Khadabadi SS; Distribution and Ancient-recent Medicinal Uses of Trichosanthes species; International Journal of Phytopharmacy; 2012; 2(4); 91-97

2. Adebooye OC; Phytoconstituents and anti-oxidant activity of the pulp of snake tomato (Tricosanthes cucumerina); African Journal of Traditional; Complimentary and Alternative Medicines; 2008; 5(2); 173-179.

3. Ojiako OA, Igwe CU, The Nutritive; Anti-Nutritive and Hepatotoxic Properties of Trichosanthes anguina (Snake Tomato) Fruits from Nigeria; Pakistan Journal of Nutrition; 2008; 7(1); 85-89.

4. Jiratchariyakul W, Frahm AW; Cucurbitacin B and dihydrocucurbilacin B from Trichosanthes cucumerina; J. Pharm. Sci.; 1992; 19(5); 12.

5. Swamy MJ, Komath SS, Nadimpalli SK; Identification of histidine residues in the sugar binding site of snake

gourd (Trichosanthes anguina) seed lectin; Biochem Mol Biol Int; 1998; 44(1); 107-116. 6. Yadava RN, Syeda Y; An isoflavone glycoside from the seeds of Trichosanthes anguina; Phytochemistry; 1994; 36(6); 1519-1521.

7. Yusuf M, Chowdhury JU, Wahab MA, BegumJ; 1994; Medicinal Plants of Bangladesh; BCSIR; Dhaka.

8. Hariti M, Rathee PS; Antibacterial activity of the unsaponifiable fraction of the fixed oil of Trichosanthes seeds; Asian journal of chemistry; 1995; 7(4); 909-911.

9. Kolte RM, Bisan VV, Jangde CR, Bhalerao AA; Anti-inflammatory activity of root tubers of Trichosanthes cucumerina (LINN) in mouse's hind paw oedema induced by carrageenan; Indian Journal of Indigeneous Medicines; 1997; 18(2); 117-21.

10. Kongtun S, Jiratchariyakul W, Mongkarndi P, Theppeang K, Sethajintanin I, Jaridasem S, Frahm AW; Thai, Cytotoxic properties of root extract and fruit juice of tricosanthes cucumerina; J Phytopharm; 1999; 6(2); 1-9.

11. Kar A, Choudhury BK, Bandyopadhyay NG; Comparative evaluation of hypoglycaemic activity of some Indian plants in alloxan diabetis rats; Journal of Ethnopharmacology; 2003; 84(1); 105-108.

12. Arawwawala M , Thabrew I, Arambewela L; Antidiabetic activity of Trichosanthes cucumerina in normal and streptozotocin–induced diabetic rats; Int. J. of Bio. and Che. Sci.; 2009; 3(2); 287-296

13. Kirana H, Srinivasan B; Tricosanthes cucumerina improves glucose tolerance and tissue glycogen in non insulin dependent diabetis mellitus induced rats; Indian Journal of Pharmacology; 2008; 40(3); 103-106.

14. Rahuman AA, Venkatesan P; Larvicidal efficacy of five cucurbitaceous plant leaf extracts against mosquito species; J. of Parasitol Res; 2008; 103(1); 133-139.

15. Kumar SS, Kumar BR. & Mohan GK; Hepatoprotective effect of Trichosanthes cucumerina Var cucumerina L. on carbon tetrachloride induced liver damage in rats; Journal of Ethnopharmacology; 2009; 123(2); 347-350

16. VanWagenen BC, R Larsen, JH Cardellina, D Randazzo, ZC Lidert, C Swithenbank; Ulosantoin, a potent insecticide from the sponge Ulosa ruetzleri; J Org Chem; 1993; 58(2); 335-337.

17. Meyer B.N., N.R. Ferrigni, J.E. Putnam, L.B. Jacobsen, D.E. Nichols and J.L. Mclaughlin; Brine shrimp: a convenient general bioassay for the active plant constituents; Planta Medicine; 1982; 45(5); 31-34.

18. Zhao, G.X., Y.-H. Hui, J.K. Rupprecht, J.L. McLaughlin, and K.V. Wood, Additional bioactive compounds and trilobacin, a novel highly cytotoxic acetogenin, from the bark of Asimina triloba; Journal of Natural Products; 1992; 55(3); 347-356.

19. Sadhu SK.; E Okuyama, H Fujimoto and M Ishibashi; Separation of Leucas aspera, a medicinal plant of Bangladesh, guided by prostaglandin inhibitory and antioxidant activities; Chemical & Pharmaceutical Bulletin 2003; 51(5); 595-598.

20. Brand-Williams W, Cuvelier ME, Berset C; Use of free radical method to evaluate antioxidant activity; Lebensm Wiss Technology; 1995; 28(1); 25-30

21. Skerget M, Kotnik P., Hadolin M., Hras AR, Simonic M, Knez, Z; Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food Chem; 2005; 89(2); 191-198.

22. Majhenic L, kerget MS, et Knez Z; Antioxidant and antimicrobial activity of guarana seed extracts; Food Chem; 2007; 104(3); 1258-1268.